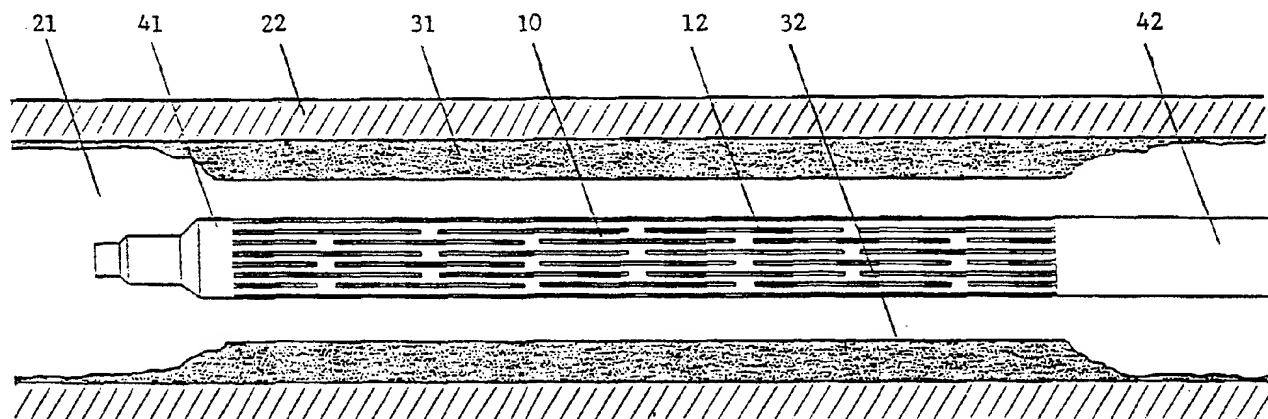




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(54) Title: MEDICAMENT DISPENSING STENTS



## (57) Abstract

A medicament dispensing medical implant is described which is fabricated from relatively non-inflammatory biogenic tissue or biopolymers for implantation in or adjacent to a target tissue in the human body. The implant, which is preferably in the form of a stent (10), is non-thrombogenic, optically translucent and relatively non-inflammatory, delivers relatively high doses of one or a combination of medicaments locally in a sustained fashion while delivering a relatively low dose of said medicaments systemically.

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## MEDICAMENT DISPENSING STENTS

## BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates to an implantable medicament dispensing stent or patch for the treatment of atherosclerosis by adjunctive photodynamic therapy generally, and more particularly to a medicament dispensing device for the prevention of restenosis following atherectomy or angioplasty.

2. Prior Art

Atherosclerosis is a cardiovascular disease in which deposits of plaques (atheromas) containing cholesterol, lipid material, foam cells, lipophages and proliferating smooth muscle cells are within the intima and media of large to small diameter arteries such as the aorta and the iliac, femoral, coronary and cerebral arteries. The resultant stenosis causes reduction in blood flow.

Attempts to treat atherosclerosis have included by-pass surgery wherein the diseased vascular segments are augmented by prosthetic or natural grafts. This procedure requires general anesthesia and a substantial healing period after surgery and, thus, is generally limited to cases of severe coronary artery disease.

Other approaches for the recanalization of stenotic vessels include percutaneous transluminal coronary angioplasty (PTCA), atherectomy, stenting and newer modalities of cardiovascular intervention including laser angioplasty. The word "recanalization", as used herein, means a procedure for increasing blood flow through the occluded vessel by angioplasty, including dilation or ablation or removal of occlusive material. The primary drawbacks of these methods has been restenosis. Studies have shown that restenosis, or the re-narrowing, of the internal lumen of an artery subsequent to such recanalization occurs in about 25-50% of cases where such primary treatment is performed. The result of re-stenosis is the requirement for an additional interventional or surgical procedure.

Various mechanisms can cause re-stenosis. One mechanism is rapid smooth muscle cell (SMC) proliferation at the lesion site. Smooth muscle cell proliferation is believed to occur immediately or at any time up to several hours after vessel wall injury that results from primary atherosclerotic treatment such as angioplasty. This proliferation continues for about 5-18

1 days depending on the individual. The cause of this rapid smooth muscle cell proliferation is  
2 believed to involve the release of various growth factors in response to the vessel wall injury.  
3 Specifically, after such vessel wall injury, some smooth muscle cells migrate to the intima where  
4 they are affected by the blood elements with which they come in contact, especially platelets and  
5 lipoproteins. Platelets contain a factor that stimulates smooth muscle cell proliferation and  
6 migration, which can result in re-stenosis.

7 March, et al., in U.S. Patent 5,116,864 provides a method for preventing re-  
8 stenosis in a patient undergoing vascular recanalization. The method comprises the systemic  
9 administration of a photoactivatable psoralen compound, preferably by an oral route, to achieve  
10 therapeutically effective serum levels. Alternatively, the psoralen may be delivered to the tissue  
11 by intra-arterial injection through a catheter. Following psoralen administration, the atheroma is  
12 illuminated with ultraviolet radiation until recanalization is complete. This is where March, et al.'s  
13 treatment ends.

14 Other drugs are used to prevent restenosis. One such drug is heparin. Heparin is  
15 an anticoagulant which when delivered systemically severely reduces the ability of the body to  
16 form blood clots. Devices for preventing restenosis such as implantable stents require massive  
17 doses of heparin which requires the patient to remain hospitalized for a long period of time.

18 Stents are presently being investigated as a treatment for cardiovascular disease.  
19 Early results indicate that stents have a similar rate of restenosis when compared to conventional  
20 interventions with the added complication of abrupt closure of the vessel. Stents have also been  
21 used as a "bail out" device to maintain the patency of a collapsing artery until another corrective  
22 medical procedure can be performed. Stents are composed of many materials including stainless  
23 steel, biodegradable polymers, collagen, gelatin, etc.

24 Local drug delivery devices such as suppositories and dermal patches have been  
25 used as indwelling devices. Indwelling devices to treat cardiovascular disease through delivery of  
26 local drugs are not clinically available.

27 One approach to the problem of sustained long term drug delivery employs  
28 implantable biodegradable polymer/drug combinations in a variety of ways to achieve a controlled  
29 regular or continuous administration of the drug. Biodegradable polymers are useful as carriers  
30 for many different types of drugs because they serve as a temporary matrix to hold the drug, but

1 do not chemically interact with the drug. As the matrix erodes, the drugs are released and can  
2 diffuse into the tissues.

3 In one prior art embodiment, a synthetic (non-biogenic) biodegradable polymer  
4 matrix is homogeneously impregnated with a medicament so that the medicament is released more  
5 or less continuously and uniformly as the supporting polymer matrix erodes. In another variation  
6 of this basic idea, a single reservoir of the drug or medicament in solution is encapsulated by a  
7 semi-porous polymer matrix. The drug diffuses continuously out of the reservoir, through the  
8 polymer, and finally to the intended delivery area. Metal stents coated with bioabsorbable  
9 synthetic polymer have also been used to deliver medicament but such metal stents are optically  
10 opaque and thrombogenic. In still a further variation, tiny discrete "pockets" of the drug are  
11 encapsulated throughout the synthetic polymer. If the polymer is biodegradable then it will  
12 completely dissolve thereby releasing all of the impregnated or encapsulated drug. The above  
13 prior art devices are known in the art and are made from synthetic polymers. The problems with  
14 implants fabricated using non-biogenic material are that such prior art implants are thrombogenic  
15 and being a "foreign body" stimulate the host's inflammatory response.

16 A device such as a medicament-dispensing stent can also be constructed from  
17 naturally occurring biopolymers and derivatives thereof or biogenic tissue. Biological materials  
18 such as bovine and porcine tissues harvested from donor animals are commonly used for  
19 implantation into the human body. They are known to be non-thrombogenic and non-inflam-  
20 matory. The porcine heart valve is one such example. Such biogenic tissues are well received  
21 and well tolerated by the host human tissues and, unlike biodegradable synthetic polymers,  
22 biogenic tissue implants are less likely to induce an inflammatory host response and are replaced  
23 over time by the host natural tissue produced *in situ*. Human tissues harvested from a human  
24 donor (autologous or heterologous) are also viable tissue types for this device.

25 Accordingly, there is a need to address the problem of smooth muscle cell  
26 proliferation in the treatment of atherosclerosis to minimize or eliminate the occurrence of  
27 restenosis.

28

29

30

## SUMMARY OF THE INVENTION

Photoatherolytic (PAL) Therapy is believed to be potentially effective for the prevention of restenosis. Photodynamic Therapy (PDT) has also been demonstrated to be very effective in the treatment of various cancers. In one embodiment, a stent-type device made from biogenic tissue and/or biopolymers is described which can be used to deliver a Photosensitive drug locally to a target tissue over a period of time. In another embodiment, a plug-type of implantable device is described which can be embedded in solid tissue thereafter to deliver medicament to a target. In still another embodiment the implant may take the form of a substantially planar patch.

In a particular preferred embodiment a medical implant is described for the local delivery of medicament to an intraluminal target tissue. The device and method involves the use of a bio-absorbable biogenic patch or stent which is impregnated or otherwise burdened with a photosensitizer drug, with or without complimentary medicaments, to locally deliver said drug(s) into target tissue on the vessel wall over a prolonged period of time as the stent is absorbed. Although the use of bioabsorbable, non-biological, but biocompatible stents have been proposed for such a therapy and remain a viable solution, these materials are often inflammatory to the host tissue.

Accordingly, it is an object of this invention to produce a medicament-dispensing implant which is non-thrombogenic, minimally inflammatory and generally well received and well tolerated by the human body.

It is another object of this invention to produce a medicament-dispensing implant which is absorbed by the body over time.

It is yet another object of this invention to produce a medicament-dispensing implant which can deliver medicaments substantially only to selected target tissues.

It is still another object of this invention to provide a biodegradable biogenic tissue implant which can deliver medicaments over a sustained period of time and be replaced by host tissue.

It is another object of this invention to describe a method in which this device can be used to locally deliver medicament(s) over a prolonged period of time.

1           It is still another object of this invention to describe a method in which this device  
2 can be used to locally deliver medicament(s) including a photosensitizer over a sustained period of  
3 time which is ultimately used in PDT.

4           The above referenced objectives are met by the present invention which is best  
5 understood by referring now to the preferred embodiments of the invention.

6           The device of the present invention provides a means for treating vascular disease  
7 which avoids the problems and disadvantages of the prior art. The invention accomplishes this  
8 goal by providing a bioabsorbable (biodegradable) stent which releases a medicament. The  
9 medicament blocks the growth factor binding sites on the atherosclerotic smooth muscle cells  
10 (SMC) injured during the recanalization until growth factor is no longer released from the  
11 platelets in the vicinity of the injured cells. In this way, smooth muscle cell proliferation in the  
12 vicinity of the lesion site is inhibited, thereby minimizing or eliminating the occurrence of  
13 restenosis.

14           The device, generally comprising of a stent, is used post recanalization to: a)  
15 deliver a photosensitive medicament which blocks the chain of events linked to restenosis along  
16 with delivering other medicaments to prevent events linked to restenosis such as thrombosis and  
17 the formation of an intravascular matrix of collagen and fibrin; b) maintain the patency of the  
18 treated artery and prevent elastic recoil of the artery by adding structure and support to the vessel  
19 wall; and c) deliver and maintain a level of photosensitizer to the treatment site which inhibits  
20 smooth muscle cell proliferation and, when activated by light energy, induces cell lysis.

21           In the preferred embodiment, the blocking step is accomplished by introducing a  
22 photosensitizing agent in the region of the vessel subject to the recanalization such that the agent  
23 accumulates in the atheromatous plaque and injured smooth muscle cells. The photosensitizer,  
24 accumulated in the atheromatous plaque and smooth muscle cells, blocks the smooth muscle cell  
25 growth factor binding sites to inhibit smooth muscle cell proliferation. The photosensitizer is  
26 slowly released as the drug-laden bioabsorbable wall of the vasoabsorbable stent (VAST) slowly  
27 disintegrates over a period of about 5-18 days following recanalization, which corresponds to the  
28 period needed for growth factor release from platelets to terminate and which varies among  
29 patients. The continuous readministration of photosensitizer serves to replace previously  
30 administered photosensitizer, which is cleared from the cells over time, to ensure that the growth

1 factor binding sites are blocked until growth factor has cleared from the tissues. After the  
2 majority of the photosensitizing agent has been delivered to the surrounding smooth muscle cells  
3 from the VAST but before it clears from the atherosclerotic smooth muscle cells and plaque, the  
4 photosensitizing agent is exposed to light at a wavelength at which the photosensitizer absorbs  
5 the light causing cell lysis. Since the growth factor has cleared before atherosclerotic plaque and  
6 cell lysis, the likelihood of restenosis is significantly reduced or eliminated. The process of  
7 activating a photosensitizer with light to cause cell necrosis is called photodynamic therapy, or  
8 more particularly in the case of atherosclerosis, photoatherolytic therapy.

9 The advantages of the at least partially degradable medicament dispensing stent  
10 over prior art include:

- 11 1. Bioabsorbable polymer;
- 12 2. Non-thrombogenic;
- 13 3. May be a coated metallic stent for reduced thrombogenicity;
- 14 4. Localized drug delivery - continuous localized time release drug delivery to  
15 affected area eliminates photosensitivity problems which arise following  
16 systemic delivery;
- 17 5. Maintain structural integrity of vessel (scaffolding);
- 18 6. Maintain patency - prevent elastic recoil;
- 19 7. Eliminate the need for serial injection of medicament;
- 20 8. Eliminate the need for serial oral administration of medicament;
- 21 9. Lower systemic doses and higher local doses of medicament;
- 22 10. Combined drug therapy (anti-proliferation, anti-recoil, anti-thrombo-  
23 genicity, anti-platelet, anti-fibrin, anti-collagen); and
- 24 11. Combined drug/device therapy.

25 In summary, the medicament-dispensing VAST of the present invention has the  
26 potential to greatly impact the treatment of cardiovascular disease by treating the disease from a  
27 cellular cause level (the atherogenesis perspective) and not merely from the conventional palliative  
28 approach.

29 Although the most important application of this novel method is to prevent  
30 restenosis following angioplasty or atherectomy of coronary arteries, this technique also can be



1 applied to atherosclerotic arteries located elsewhere, such as the renal, iliac, femoral and popliteal  
2 arteries. Additionally, a vaso-absorbable stent can be used to prevent arterial occlusion after  
3 coronary by-pass surgery wherein vascular segments are replaced with prosthetic or natural grafts  
4 and growth factor is released in response to the arterial wall injury.

5 The above is a brief description of some deficiencies in the prior art and ad-  
6 vantages of the present invention. Other features, advantages and embodiments of the invention  
7 will be apparent to those skilled in the art from the following description, accompanying drawings  
8 and appended claims.

9

#### 10 BRIEF DESCRIPTION OF THE DRAWINGS

11 Figure 1 depicts a stent in its non-expanded state.

12 Figure 2 (a,b,c) depicts an artery with an atheromatous plaque.

13 Figure 3 (a,b) depicts the same artery as Figure 2(a-c), but with views before and  
14 after an atherectomy procedure has been performed to removed the atheromatous plaque.

15 Figure 4 depicts the stent loaded over and angioplasty balloon in a non-expanded  
16 state at the lesion site post atherectomy.

17 Figure 5 depicts the stent expanded over the expanded angioplasty balloon at the  
18 lesion site post atherectomy.

19 Figure 6 depicts the stent deployed in the artery post atherectomy where it remains  
20 until it is absorbed by the artery wall thus locally delivering a cocktail of medicament continuously  
21 to the arterial wall.

22 Figure 7 is a partially cutaway perspective view of the stent deployed in an artery.

23 Figure 8 is a side view of a medicament-dispensing implant in accordance with the  
24 present invention in the form of a stent.

25 Figure 9 shows the stent of Figure 8 deployed on a balloon catheter.

26 Figure 10 is a cut-away view of a tubular tissue such as an artery housing the  
27 catheter of Figure 9 with the balloon only partially inflated.

28 Figure 11 is a cut-away view of a tubular tissue such as an artery housing the  
29 catheter of Figure 9 with the balloon fully inflated.

1                   Figure 12 shows a partially cut-away view of the artery with the stent deployed in  
2 the artery adjacent to the target tissue and with the catheter removed.

3                   Figure 13 is a partially cutaway perspective view of the artery showing the stent  
4 deployed within the artery as in Figure 12.

5                   Figure 14 is a partially cutaway perspective view of the shaft of a needle showing  
6 an implantable plug which is dimensioned to fit within the lumen of the hollow-bore needle for  
7 deployment.

8

## 9                   DESCRIPTION OF THE PREFERRED EMBODIMENT

### 10                   Preparation of the Biogenic Tissue

11                   Biogenic tissue such as endothelium from the innermost layer of an artery (intima),  
12 collagen, fibrin, etc. is surgically removed from a donor animal such as swine or a human donor  
13 (autologous or heterologous) and maintained in a nutrient rich solution. These viable tissues can  
14 then be "fixed" using a stabilized glutaraldehyde process at pressures less than 2mm Hg that is  
15 well known in the art and used for such implants as replacement porcine heart valves.

16                   Alternatively, biogenic macromolecules (alternatively referred herein as  
17 "biopolymers"), such as a collagen, chitin, chitosan or cellulose may also be used to fabricate a  
18 biogenic implant. Chitin, for example, comprises the bulk of the organic material in arthropod  
19 exoskeletons such as crab shell. If the exoskeleton is demineralized using strong acid the  
20 remaining chitinous fraction may be extracted, deacetylated (if desired), and pressed in to the  
21 desired shape for implantation. Cellulose is a polymer comprising glucose rings derived from  
22 plants. Derivatives of cellulose which may be suitable for implantation include methylcellulose,  
23 ethylcellulose, hydroxyethylcellulose, hydroxypropylcellulose and sodium carboxymethylcellulose.

24

### 25                   Preparation of the Biogenic Implant

26                   After the biogenic tissue has been prepared as described above, the biogenic tissue  
27 is soaked in a solution containing a relatively high concentration of the desired medicament(s)  
28 such as, for example, a photosensitizer (i.e. 0.1 - 1.0 mg of PS/ml solution - . . . the desired  
29 concentration of PS in the device implantable will depend on the thickness of the lesion being  
30 treated and its location) for a period of 1-5 hours (the time will again depend on the tissue

1 absorption characteristics and the location and application of the implant). The biogenic tissue is  
2 maintained at 37 ° C during the PS drug-burdening process while excluding light at wavelengths  
3 which might activate the PS. Other medicaments may be added to the bath permeate the implant.  
4 One such drug which is potentially complimentary to the PS is heparin which has the  
5 characteristics being an anti-coagulant, anti-platelet, anti-fibrin, anti-collagen agent and may  
6 facilitate the prevention of restenosis. New molecules having Heparin-like activity may also be  
7 employed.

#### 8 Method of Using the Biogenic Implant

9 An exemplary method for using the biogenic implant according to the present  
10 invention may taught by looking first at a stent as shown in Figure 8. The stent 80 is a tubular  
11 member comprising a biogenic tissue 81 with medicament 82 incorporated therein. The  
12 medicament may be a photosensitizer which has been shown to be useful for preventing restenosis  
13 following atherectomy, or it may be any other medicament which is desirable to have released  
14 over a long period of time. The medicament may be encapsulated in discrete cells or evenly  
15 distributed throughout the body of the stent.

16 Turning now to Figure 9 we see the stent 80 with a balloon catheter 92 having a  
17 distal tip 93 inserted therein. The balloon catheter 92 has a balloon portion 91 which can be  
18 partially inflated so that the outer surface of the balloon portion 91 firmly and snugly engages the  
19 inner surface of the stent 80. A sheath (not shown) may or may not be used over the  
20 balloon/stent catheter to prevent the undesired deployment of the stent while advancing and  
21 positioning the catheter. Standard angioplasty procedures are used to deliver the stent through  
22 the femoral artery or other arterial point of entry and advancing the stent to the site of the target  
23 tissue.

24 In Figure 10, a longitudinal, cross-sectional view of an arterial member is shown  
25 having an arterial wall 102 and a lumen 101. A atheromatous patch 103 on the wall 102 of the  
26 vessel (the "target tissue" in this example) has been partially removed to permit passage of the  
27 catheter 92 therethrough. The catheter 92 is advanced through the vessel until the balloon  
28 portion 91 directly underlies the area of the vessel in which it is desirable to deploy the stent 80.  
29 Once in position, the stent can be deployed by inflating the balloon member 91 so that the outer  
30 surface of the stent 80 pressed against the atheromatous lesion 101 on the wall of the vessel 102

1 as shown in Figure 11. The expanded stent may then be "welded" to the atheromatous tissue 103  
2 on the wall 102 of the vessel. This "welding" may be accomplished by the application of heat to  
3 the stent. Once the stent 80 has been deployed and adhered to the target tissue 101 and the wall  
4 of the vessel, the balloon portion 91 of the catheter 92 is deflated and catheter removed as shown  
5 in Figure 12. In Figure 13 we see a fragmentary section of the vessel wall with the stent deployed  
6 therein and the balloon catheter removed. With the foregoing description in mind, I present an  
7 example of the device and method used to prevent restenosis following atherectomy.

8 According the present invention, photodynamic therapy is used as an adjunctive  
9 procedure to primary atherosclerotic treatment, such as percutaneous transluminal coronary  
10 angioplasty, laser angioplasty, and atherectomy to minimize or eliminate the occurrence of  
11 restenosis. Photodynamic therapy is more appropriately called photoatherolytic therapy (light  
12 induced atheromatous SMC lysis) in the specific case of cardiovascular disease.

13 Photodynamic therapy involves the administration of a particular medicament  
14 called a photosensitizer, usually by intravenous injection into an atherosclerotic patient. The  
15 photosensitizer, when administered intravenously, is selectively retained by the atheromatous  
16 smooth muscle cells and plaque, with little or no retention into healthy areas of the arterial wall.  
17 Generally, the photosensitizer is nontoxic to all cells when administered, but once activated by a  
18 therapeutic dose of light commonly delivered by a laser at a specific wavelength, the  
19 photosensitizer, which has been selectively absorbed in the atherosclerotic cells, becomes toxic.  
20 In this way, the activated photosensitizer facilitates the destruction and reabsorption of the host  
21 atheromatous plaque and smooth muscle cells (cell necrosis). The mechanism of cell necrosis  
22 induced by photodynamic therapy is believed to involve a photochemical reaction that produces a  
23 species of oxygen called singlet oxygen, which induces cell death.

24 Since the surrounding healthy tissue does not retain the photosensitizer to the  
25 extent the diseased tissue does, the therapeutic dose of light is benign to the healthy tissue regions  
26 resulting in selective necrosis. This process is disclosed using Hematoporphyrin Derivative  
27 (HPD) as the photosensitizer in U.S. Patent No. 4,512,762 to Spears, the disclosure of which is  
28 hereby incorporated herein by reference.

29 According to the present invention, the photosensitizer is administered in such a  
30 way as to inhibit smooth muscle cell proliferation following recanalization. It has been found that

1 photosensitizers that have accumulated in smooth muscle cells act in the manner of a competitive  
2 inhibitor to block the growth factor binding site, thus preventing the smooth muscle cells from  
3 getting "switched on" by growth factor, which would otherwise cause rapid cell proliferation.  
4 However, since proliferation of smooth muscle cells occurs immediately or at any time up to  
5 several hours after vessel wall injury and continues for about 5 to 18 days (depending on the  
6 individual), the timing of the administration of the photosensitizer is critical to the present  
7 invention. The effect of the timing of the photosensitizer administration is discussed in detail  
8 below.

9               Growth factor is released in response to arterial wall injury as a result of the  
10 primary treatment (e.g., angioplasty). Since release of growth factor continues for about 5-18  
11 days after arterial wall injury, the ubiquitous growth factors are free to "switch on" the remaining  
12 smooth muscle cells, resulting in rapid smooth muscle cell proliferation and restenosis. Thus,  
13 although concurrent or sequential recanalization with coronary angioplasty (or other  
14 interventional therapy) and photodynamic therapy (using a single administration of  
15 photosensitizer) reduces the initial proliferation of smooth muscle cells, in the long term such  
16 treatment may not be effective.

17               According to the present invention the preferred method of administering  
18 photosensitizer and providing adjunct therapy to cardiovascular intervention is by means of a  
19 vaso-absorbable, medicament dispensing stent. Turning now to the drawings, the "base material"  
20 of the stents 10 and 80 shown in Figure 1 and Figure 8 respectively, is preferably a polymer which  
21 has the characteristic of being absorbed by the vessel within 30-180 days. Suitable polymers or  
22 copolymers include polyglycolic/polylactic acid (PGLA), polycaprolactone (PCL),  
23 polyhydroxybutyrate valerate (PHBV) and the like. Since this therapy requires a series of  
24 pharmaceuticals to be delivered at specific times, the VAST device 10 is preferably produced in  
25 layers, each layer dispensing photosensitizer concentrations to maintain the required levels in  
26 adjacent atheromatous tissue. The stent 10 can also be made from stainless steel, titanium,  
27 NITINOL, or a variety of other metals. The drugs can be deposited on the metallic stent in a  
28 dispensing coating layer. The thickness of the drug layer 12 will depend on the time interval in  
29 which the drug is desired to be delivered (e.g. 30 days).

1           If the VAST 10 or 80 is made from a bio-absorbable polymer 11, the photo-  
2 sensitive medicament 12 will be impregnated throughout the VAST device to maintain the  
3 therapeutic levels during disintegration of the polymer. Photosensitive medicament acts as an  
4 inhibitor of smooth muscle cell (SMC) proliferation following vascular injury while also mediating  
5 SMC lysis when activated with light energy. One such photosensitive medicament is Tin Ethyl  
6 Etiopurpurin (SnET2). SnET2 has been shown to be selectively retained by atheromatous  
7 plaques. Table 1 is a list of potentially useful photosensitive medicaments.

8           Preferably, an anti-platelet/anti-thrombus drug such Heparin, Hirudin, tPA,  
9 Streptokinase, Urokinase, Persantine, Aspirin, etc. impregnates the VAST. Since restenosis is a  
10 response to injury, platelets are deposited at the lesion site. Platelets, along with other cellular  
11 components, release growth factors such as Platelet Derived Growth Factor (PDGF) which  
12 activates SMC proliferation. Reducing the number of platelets which get deposited at the site of  
13 injury and reducing the incidence of thrombus at the lesion site will greatly enhance the  
14 procedure. Thrombus has been associated with abrupt reclosure, embolism and the genesis of  
15 intravascular matrix which later becomes fibrous in advanced lesions.

16           After approximately 14 days, the SMC proliferation is replaced by the aggregation  
17 of fibrin and collagen as the dominant event in restenosis. An anti-collagen/anti-fibrin drug such  
18 as Heparin is staged to be released after about 14 days.

19           By taking this combined pharmacological regime, the proliferative events can be  
20 prevented while the normal healing process occurs. Once the healing has occurred and the  
21 growth factors have cleared from the lesion site, the light activation of the photosensitizer can  
22 occur if necessary.

23           Clinical Method: A patient presents himself/herself to the cardiac catheterization  
24 laboratory with a 90% stenosis in the mid left anterior descending (LAD) coronary artery. It is  
25 decided that the patient will be treated with an atherectomy procedure to remove a large portion  
26 of the obstruction followed by Photo-atherolytic (PAL) Therapy to prevent restenosis.

27           The diseased artery is shown in Figures 2a-c. The diseased artery, a segment of  
28 which is shown at 20 in Figure 2a, has an at least partially open central lumen 21 and an arterial  
29 wall 22. The wall 22 is comprised of concentric layers of tissue. The outermost layer is the  
30 adventitia 23. The external elastic lamina 24 separates the adventitia 23 from the media 25. The

1 internal elastic lamina 26 forms the boundary between the media 25 and the intima 27. The  
2 segment of diseased vessel 20 is shown in the longitudinal cross section in Figure 2(b). The  
3 occluded portion comprising the atherosclerotic plaque is enlarged and shown in greater detail in  
4 Figure 2(c). The atheroma consists of a necrotic core 28 surrounded by a layer of smooth muscle  
5 cells 29.

6 The patient is prepared per normal angioplasty procedures including the proper  
7 level of anti-coagulation prior to the atherectomy procedure. Turning now to Figure 3, the  
8 atheroma 31 is removed according to standard procedures. Following the atherectomy procedure  
9 the majority of the atheroma 31 (Figure 3(a)) has been removed as shown at 32 in Figure 3(b)  
10 leaving only a small amount of residual atheromatous tissue.

11 Following atherectomy as outlined above, the VAST device 10 or 80 is introduced  
12 into the vessel over a standard angioplasty balloon catheter 42 as shown in Figure 4. Once at the  
13 site of the lesion the VAST 10 is deployed by inflating the balloon portion 42 of the catheter 41 as  
14 shown in Figure 5. The balloon catheter 42 is then removed leaving the VAST in place as shown  
15 in Figures 6 and 7. The patient is then allowed to recover as with a standard atherectomy  
16 procedure. Since the VAST contains Heparin or another anti-coagulant, heavy anti-coagulation is  
17 not required as with the placement of standard metallic stents. It is advisable to prescribe an  
18 aspirin per day for the duration of the therapy. During that time the VAST releases the cocktail of  
19 therapeutic agents to the residual atheromatous lesion in a time release fashion.

20 Approximately 30 days following placement of the VAST, the patient presents to  
21 the cardiac cath lab. A sample of the patient's blood is taken and tested for levels of  
22 photosensitizer, anti-coagulant/anti-thrombus agent and anti-collagen/anti-fibrin agent. The level  
23 of photosensitizer (PS) is observed closely to determine if a systemic injection of the PS is  
24 required to effectively perform the Photodynamic Therapy (PDT) part of the procedure. If  
25 additional PS is required, the typical delay time for optimum photosensitization (i.e., 24 hours)  
26 must be observed.

27 If smooth muscle cell proliferation is controlled early enough and control  
28 maintained until growth factors clears, and if the arterial lumen is sufficiently widened, the light  
29 activation therapy may not be necessary, making this solely a pharmacokinetic therapy.

Photosensitizers which may be dispensed by the stent according to this invention include the following classes: purpurins, verdins, chlorins, phthalocyanines, phorbides, bacteriochlorophylls, porphyrins, chalcogenapyryliums, texaphyrins, xanthenes, benzophenoxazines, phenothiazines, di- and triaryl-methanes, and kryptocyanines. Preferred members of the above classes are listed in the following table. The optimum light wavelength for activating each member to achieve necrosis is provided in the right column.

TABLE 1

<u>CLASS</u>	<u>PREFERRED COMPOUND</u>	<u>ACTIVATION Wavelength (nm)</u>
Purpurins (metalized)	Tin Ethyl Etiopurpurin	660
	Ethyl Etiopurpurin	695
Purpurins (non-metalized)	Coproverdin-II-Tripotassium	700
Verdins	Salt	650
Chlorins	Octaethyl	
Phthalocyanines	Chloaluminum Sulfonated	665
	Phthyalocyanine	660
Phorbides	Mono-L-Aspoaryl Chlorin e6	780
Bacteriochlorophylls	Bacterochlorophyll-a	630
Porphyrins	Protoporphyrin-IX	800
Chalcogenapyryliums	Chalcogenapyrylium 8b	780
Texaphyrins	Texaphyrin	480-520
Xanthenes	Rhodamine 123	680
Benzophenoxazines	Nile Blue	660
Phenothiazines	Methylene Blue	660
Di and Triaryl Methanes	Victoria Blue-BO	660-700
Kryptocyanines	EDKC*	

\*EDKC = N, N-bis[2 ethyl-1, 3-dioxolane] kryptocyanine

The preferred photosensitizing agent for incorporation in and dispensation by the VAST is Tin Ethyl Eitopurpurin having the chemical name: Ethyl 3,4,20,21-tetradehydro-4,9,14,19-tetraethyl-18,19-dihydro-3,8,13,18-tetramethyl-20-phorbine carboxylato(2-)-N<sup>23</sup>, N<sup>24</sup>, N<sup>25</sup>, N<sup>26</sup> - tin(IV) dichloride. It has been found that there is little or no retention of this drug in the skin, thereby avoiding problems that can result from exposure of the patient to ordinary



1 sunlight (i.e., activation of the photosensitizer in the skin). In addition, Tin Ethyl Etiopurpurin  
2 has a high therapeutic ratio (concentration level in diseased tissue relative to healthy tissue) as  
3 compared to other photosensitizers such as hematoporphyrin derivative (HPD). Tin Ethyl  
4 Etiopurpurin also is advantageously activated at longer wavelengths (660-690 nanometers). At  
5 these wavelengths, the light is attenuated to a much lesser degree by the blood as is the case with  
6 630 nanometer wavelength light (the optimum wavelength for HPD, for example). As a result of  
7 using a longer activation wavelength, a substantially greater amount of light gets to the vessel  
8 wall and photosensitizer, thereby increasing procedure efficiencies.

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10 Example of Using the Biogenic Stent for Preventing Restenosis

11 Following Atherectomy or Angioplasty

12 A drug-burdened biogenic implant in the tubular form of a stent 80 is loaded over  
13 a light diffusing catheter such as described by Narciso, Jr. in U.S. patent #5,169,395. A sheath  
14 may or may not be used over the balloon/stent catheter to prevent the undesired deployment of  
15 the stent while advancing and positioning the catheter.

16 To deploy the stent 80, standard angioplasty procedures are used to deliver the  
17 stent through the femoral artery or other arterial point of entry. In summary, following  
18 identification of the target tissue comprising atheromatous plaque on the wall of the vessel, the  
19 stent should be deployed in the area of injury and utilized according to the following steps:

- 20 a. the stent-deploying balloon catheter is positioned at the lesion site imme-  
21 diately after the completion of the angioplasty/atherectomy (the protective  
22 sheath should be pulled back to expose the PS-laden stent at this point if a  
23 sheath was used);
- 24 b. the balloon is expanded until the surface of the PS-laden biogenic stent  
25 implant fully engages the arterial wall for the full 360 °;
- 26 c. the tissue is irradiated through the transparent balloon wall with a wave-  
27 length of light (i.e. 800-1000 nm) which produces low level heating to  
28 cause the denaturation and "welding" of the biogenic stent to the host  
29 vessel. The wavelength of the light used should not be one which activates  
30 the PS in the stent. If the activation wavelength and the welding

wavelength overlap, an alternative method of heating should be employed. As example of an alternative heating method, a radio frequency (RF) heated balloon or a balloon which incorporates circulating hot fluid may be employed to effect "welding". Alternatively, a photochemical cross-linking dye may be employed to facilitate the welding process. Such dyes include, for example, brominated 1, 8-naphthalimide compounds. These dyes are activated by visible light and, following activation, covalently bind to amino acid residues, both free and in proteins, rendering them useful as protein and peptide cross-linking agents. Care should be taken to choose a photochemical cross-linking dye with an activation wavelength which will not activate the PS if one is present. The absorption maximum for the naphthalimide compounds is around 420nm, well removed from the activation wavelength of PS compounds used in PDT.

- d. the balloon is then deflated and all catheters are removed from the body; and
- e. (Only used for PDT applications.) A predetermined time later, the PS which has been absorbed by the vessel should be activated by means of a suitable light delivery catheter such as the catheter described by Narciso, Jr. in U.S. Patent #5,169,395 and using standard PAL Therapy techniques. The PS acts to inhibit the proliferation of cells post angioplasty thereby reducing restenosis.

The foregoing procedure for preventing restenosis is exemplary and not limiting. Similar methods can be employed to deploy a stent in non-arterial lumens such as the colon or esophagus for PDT treatment of cancer. Once deployed in the artery or other tubular tissue of the host, the stent will locally deliver the medicament(s) to the lesion area over a sustained period of time (i.e. 2 weeks to 6 months). Two weeks post stent placement, re-endothelialization should occur covering the stent with natural autologous endothelium thus encapsulating the stent in the luminal wall.

If necessary, two weeks to six months following the deployment of the stent, the patient may be brought back to the catheterization laboratory. Using standard angioplasty

1 techniques and an invasive intravascular light diffusing catheter, the lesion site can be irradiated to  
2 receive a dose of light sufficient to activate the PS causing cell lysis and cell necrosis.

3           The medicament-dispensing biogenic implant of the present invention may be  
4 formed into a patch or plug for insertion into a target tissue such as a solid tumor. Such a  
5 plug 140 is shown in Figure 14. The plug 140 is dimensioned to fit within the bore of a needle  
6 141 having a tip 142. The needle 141 may be inserted into the target tissue until the tip is  
7 embedded within or adjacent to the target tissue. The plug 140 may then be extruded through the  
8 tip 142 and the needle 141 removed.

9           The above is a detailed description of particular embodiments of the invention. It  
10 is recognized that departures from the disclosed embodiment may be made within the scope of the  
11 invention and that obvious modifications will occur to a person skilled in the art. The full scope  
12 of the invention is set out in the claims that follow and their equivalents. Accordingly, the claims  
13 and specification should not be construed to unduly narrow the full scope of protection to which  
14 the invention is entitled.

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## CLAIMS

What I claim is:

1. A stent dimensioned to fit within a blood vessel wherein at least a portion of said stent is bioabsorbable and wherein at least a portion of said bioabsorbable portion comprises at least one medicament.

2. The stent of Claim 1 wherein at least one said medicament is a smooth muscle cell proliferation inhibitor.

3. The stent of Claim 2 wherein said medicament is photosensitive.

4. The stent according to Claim 1 wherein at least one said medicament is thrombolytic.

5. The stent according to Claim 1 wherein at least one said medicament is fibrinolytic.

6. A stent dimensioned to fit within a blood vessel wherein at least a portion of said stent is bioabsorbable and wherein at least a portion of said bioabsorbable portion of said stent comprises a cocktail consisting of a plurality of medicaments.

7. The stent according to Claim 6 wherein said cocktail comprises, in combination, two or more of the medicaments selected from the following group:

(a) smooth-celled proliferation inhibitors,

(b) anti-thrombotic compounds,

(c) anti-fibrin compounds,

(d) anti-collagen compounds.

8. A medicament-dispensing medical implant comprising a biocompatible bioabsorbable biogenic material impregnated with at least one medicament.

9. The medical implant of Claim 8 wherein said biogenic material comprises at least a portion of an exogenous organ.

10. The medical implant of Claim 8 wherein said biogenic material comprises at least a portion of an endogenous organ.

11. The medical implant of Claim 9 wherein said at least a portion comprises endothelium.

- 1           12.    The medical implant of Claim 10 wherein said at least a portion comprises  
2    endothelium.
- 3           13.    The medical implant of Claim 8 wherein said biogenic material comprises a  
4    biopolymer.
- 5           14.    The medical implant of Claim 13 wherein said biopolymer is cellulose.
- 6           15.    The medical implant of Claim 16 wherein said biopolymer is methylcellulose.
- 7           16.    The medical implant of Claim 13 wherein said biopolymer is collagen or  
8    derivatives thereof.
- 9           17.    The medical implant of Claim 13 wherein said biopolymer is chitin or chitosan.
- 10          18.    The medical implant of Claim 8 wherein said biogenic material is formed into a  
11   tubular stent.
- 12          19.    The medical implant of Claim 8 wherein said biogenic material is formed into a  
13   plug.
- 14          20.    The medical implant of Claim 8 wherein said at least one medicament is pho-  
15   tosensitive.

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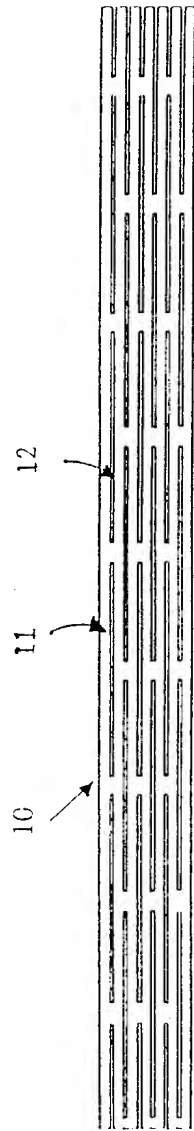


Figure 1

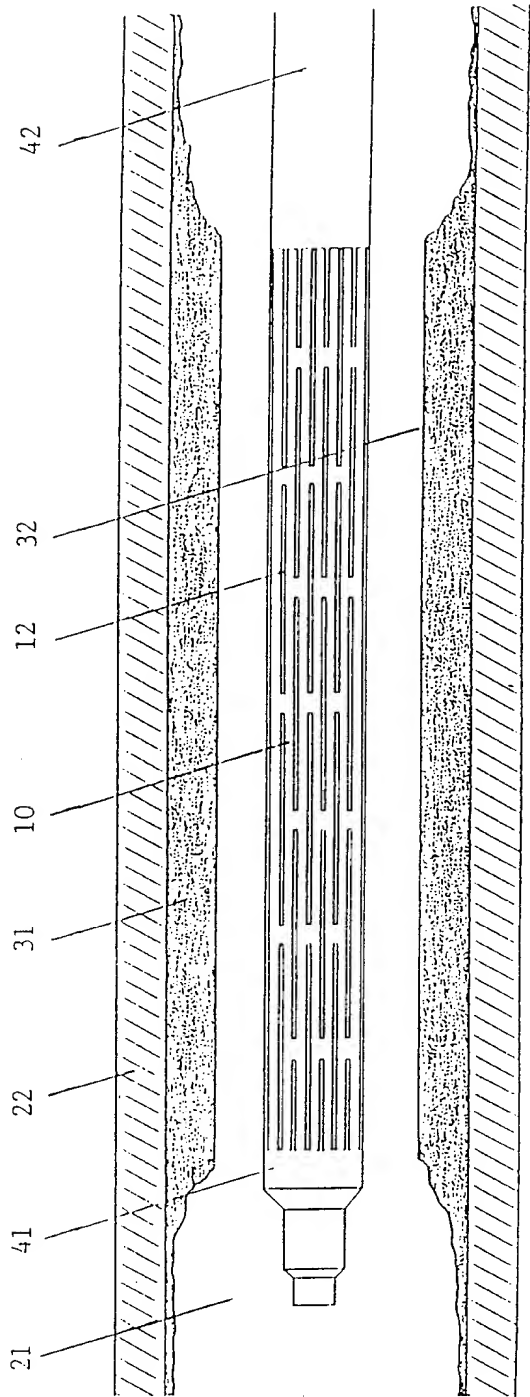


Figure 4

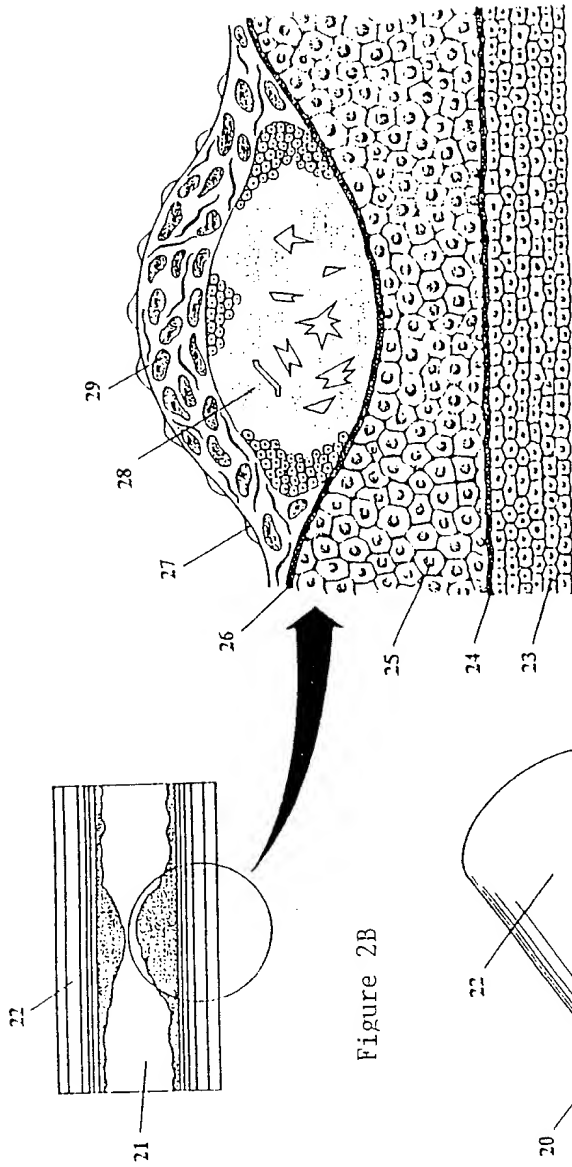


Figure 2B

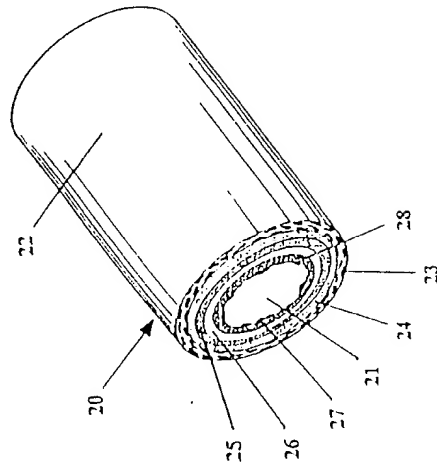


Figure 2A

Figure 2C

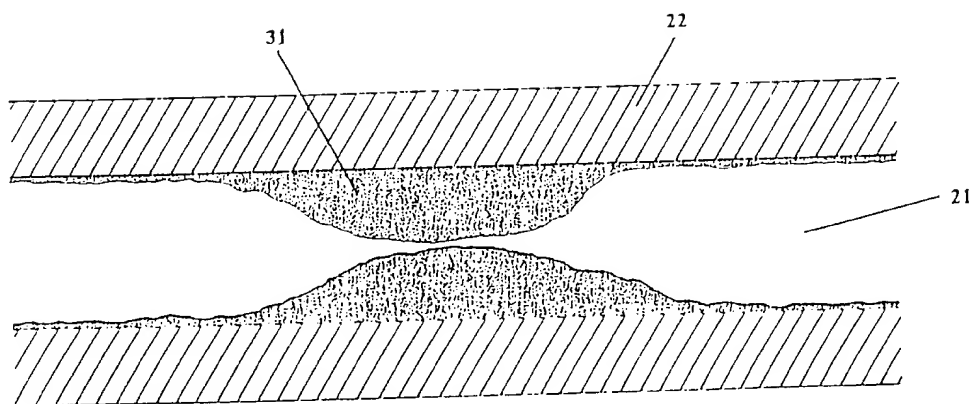


Figure 3A

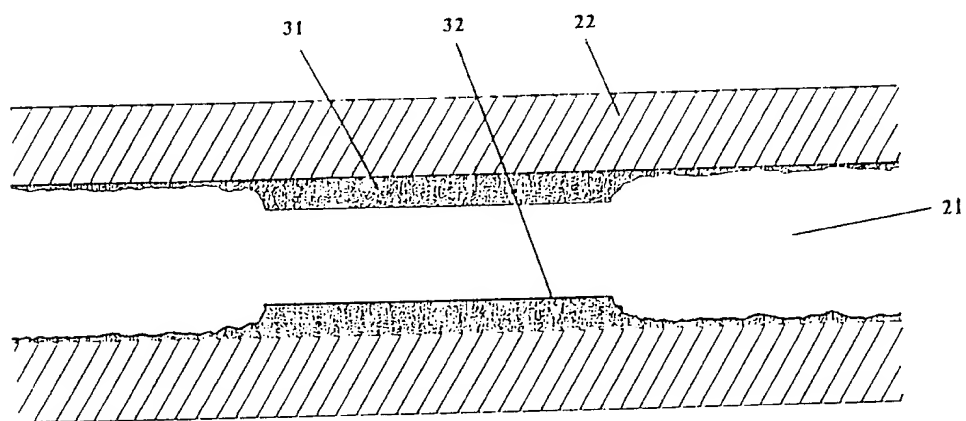


Figure 3B



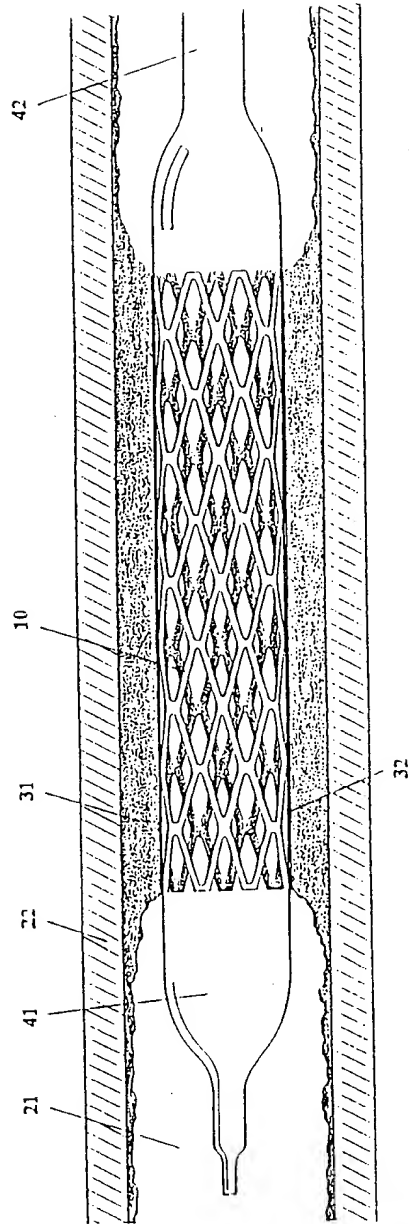


Figure 5

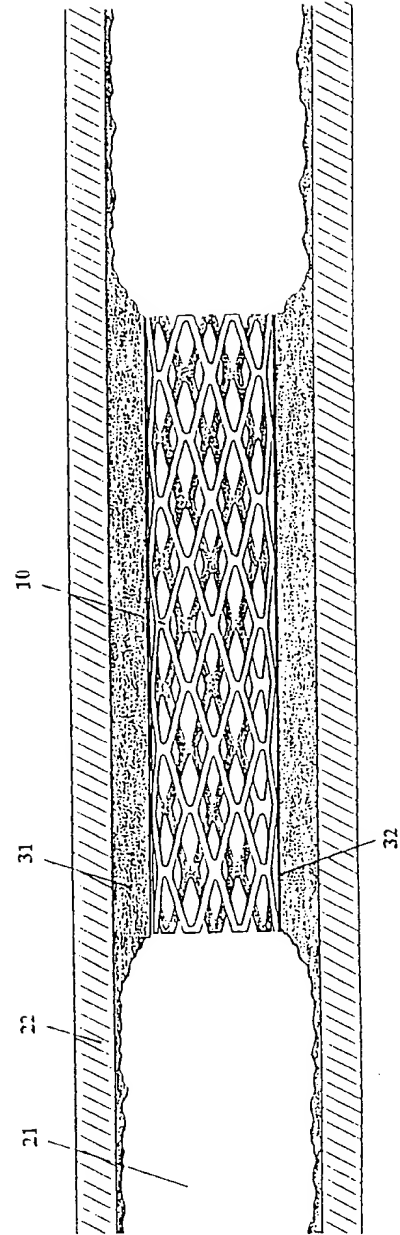


Figure 6

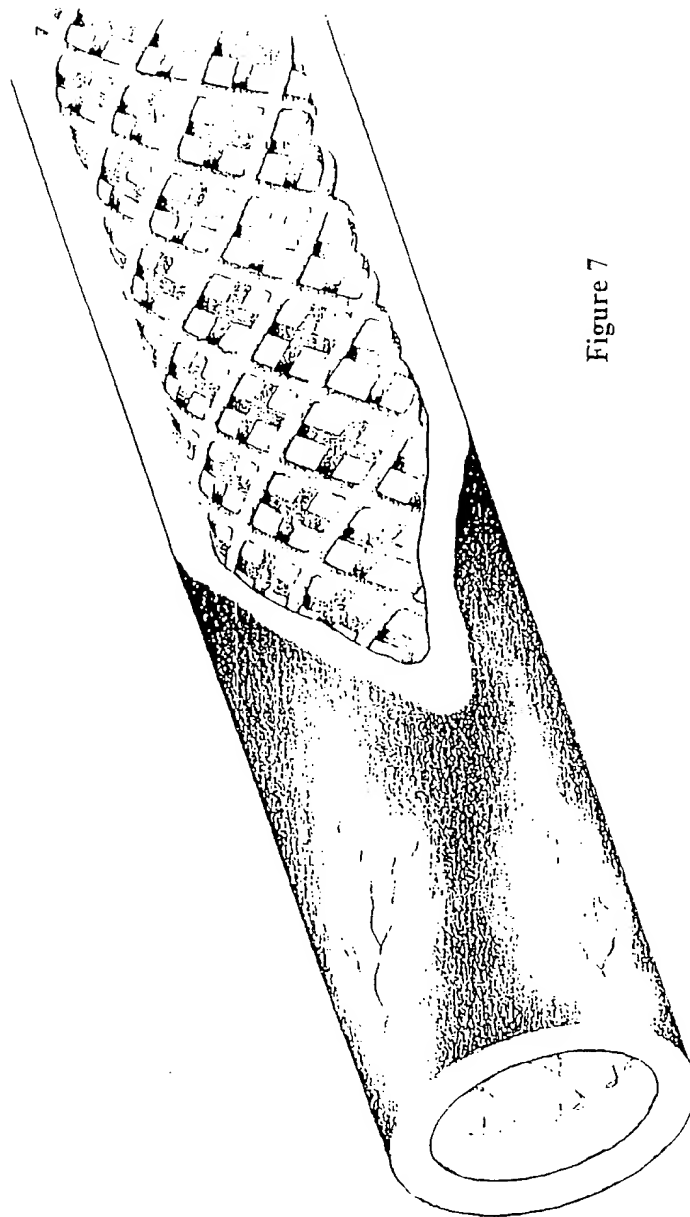


Figure 7

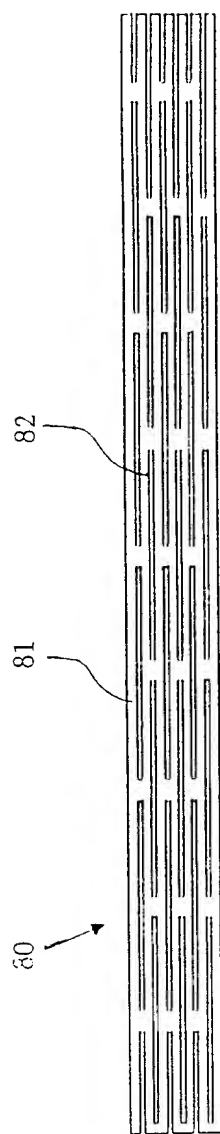


Figure 8

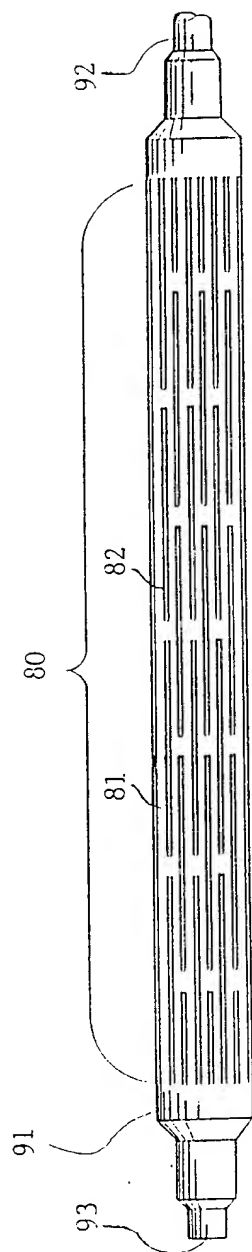


Figure 9

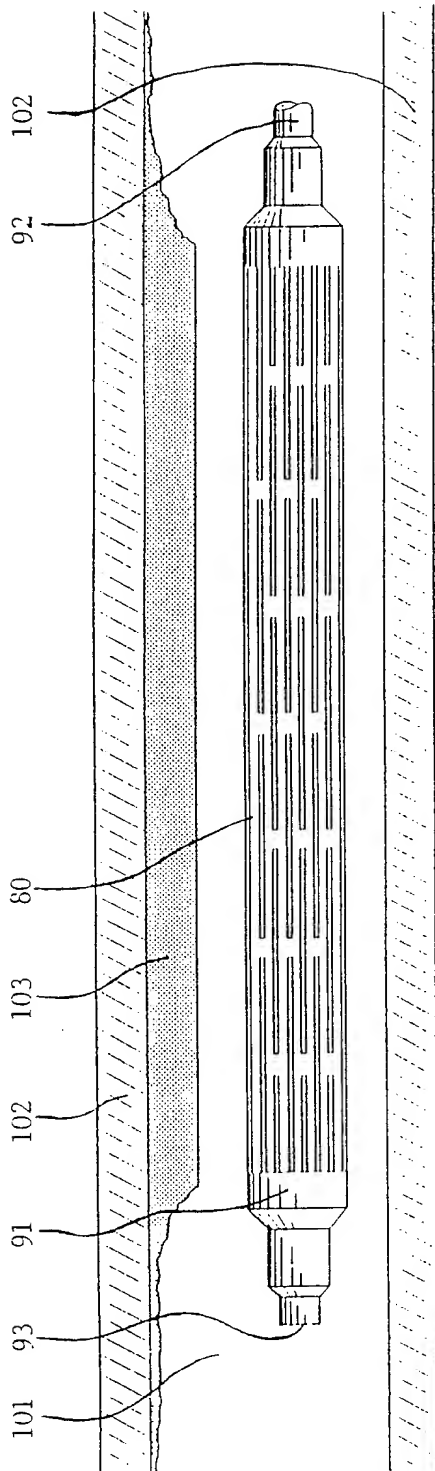


Figure 10

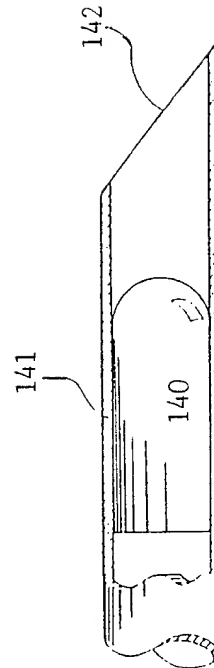


Figure 14

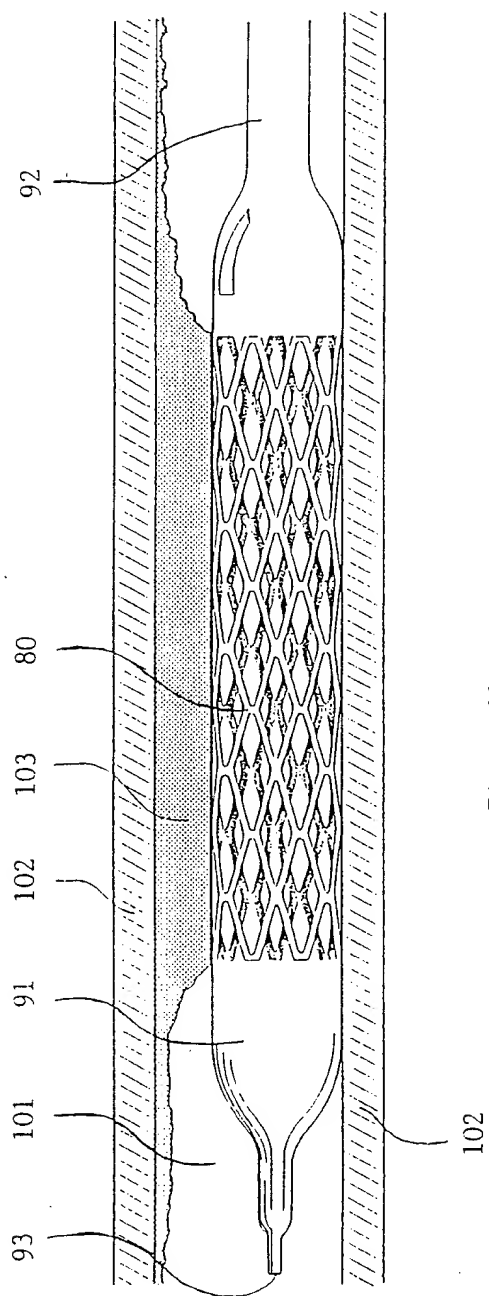


Figure 11

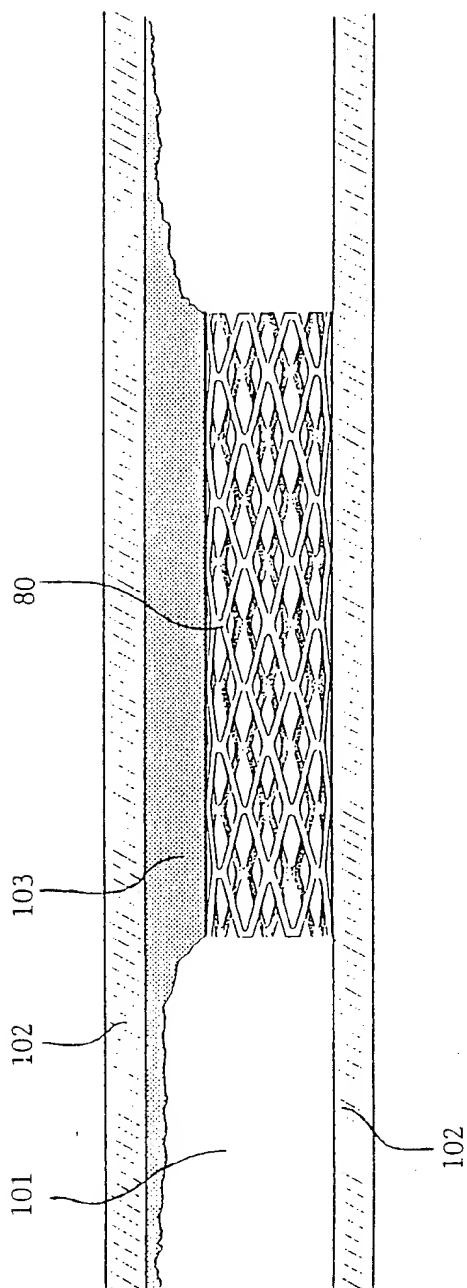


Figure 12

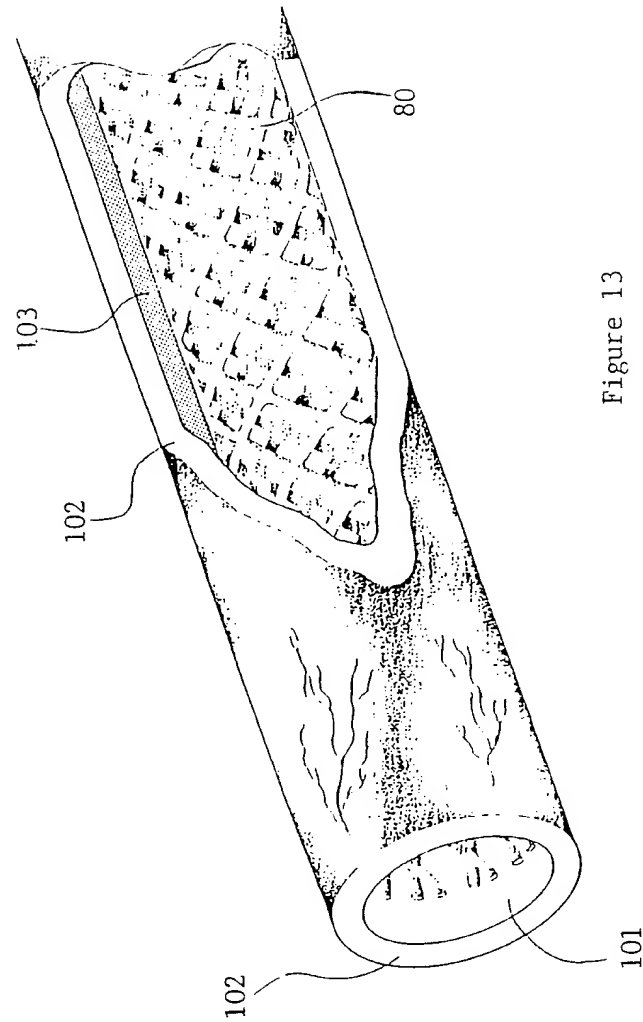


Figure 13

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US94/00369

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :A61K 9/00

US CL :604/194, 891.1

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 604/8-10, 21, 194, 890.1, 891.1; 606/154, 194; 623/1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched  
NONE

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
NONE

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X ---, P Y	US, A, 5,234,456, (SILVESTRINI), 10 August 1993. See entire reference.	1, 4 ----- 2, 3, 5-7
Y	US, A, 5,019,090, (PINCHUK), 28 May 1991, See entire reference.	5
Y, P	US, A, 5,197,977, (HOFFMAN, JR. ET AL.), 30 March 1993. See entire reference.	6, 7
Y	US, A, 5,019,075, (SPEARS ET AL.), 28 May 1991. See entire reference.	2, 3
X --- Y	US, A, 4,895,724, (CARDINAL ET AL.), 23 June 1990. See entire reference.	1, 6, 10-12 ----- 13

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A* document defining the general state of the art which is not considered to be part of particular relevance	*X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G*	document member of the same patent family
*O* document referring to an oral disclosure, use, exhibition or other means		
*P* document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

12 MARCH 1994

Date of mailing of the international search report

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## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US94/00369

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US, A, 4,849,226, (GALE), 18 July 1989. See entire reference.	7. 8
X, P	US, A, 5,264,214, (RHEE ET AL.), 23 November 1993. See entire reference.	9